1	Influence of high-protein and high-carbohydrate diets on serum lipid and fructosamine
2	concentrations in healthy cats
3	
4	Chad F. Berman ^{1,2} , Remo G, Lobetti ¹ , Eric Zini ^{3,4,5} , Geoffrey T. Fosgate ⁶ , Johan P. Schoeman ²
5	
6	¹ Bryanston Veterinary Hospital, Bryanston, Johannesburg, South Africa
7	
8	² Department of Companion Animal Clinical Studies, University of Pretoria, South Africa
9	
10	³ Clinic for Small Animal Internal Medicine, Vetsuisse Faculty, University of Zurich, Switzerland
11	
12	⁴ Department of Animal Medicine, Production and Health, University of Padova, Italy
13	
14	⁵ Istituto Veterinario di Novara, Granozzo con Monticello, Italy
15	
16	⁶ Department of Production Animal Studies, University of Pretoria, South Africa
17	
18	Corresponding author
19	Chad Farryl Berman
20	C_berman@hotmail.com
21	PO Box 67092, Bryanston 2021, South Africa
22	Key words: High-protein; high-carbohydrate; lipid profile; fructosamine

24 Abstract

25

Objectives: The aim of this study was to determine whether high-protein and high-carbohydrate diets exert
 differential effects on serum cholesterol, triglyceride and fructosamine concentrations in healthy cats.

Methods: A randomised, crossover diet trial was performed in thirty-five healthy shelter cats. Following baseline health assessments, cats were randomised into groups receiving either a high-protein or high-carbohydrate diet for four weeks. The cats were then fed a washout diet for four weeks before being transitioned to whichever of the two studied diets they had not yet received. Fasting serum cholesterol, triglyceride and fructosamine concentrations were determined at the end of each four-week diet period.

Results: Cats on the high-carbohydrate diet had significantly lower serum cholesterol (P<0.001) concentrations compared to baseline measurements. Cats on the high-protein diet had significantly higher serum cholesterol (P<0.001) and triglyceride (P<0.001) concentrations, yet lower fructosamine (P<0.001) concentrations compared to baseline measurements. In contrast, overweight cats (body condition score >5) had lower cholesterol (P=0.007) and triglyceride (P=0.032) concentrations on the high-protein diet than cats within other body condition score groups.

39 Conclusions and relevance: Diets higher in protein and lower in carbohydrates appear beneficial for short-40 term glucose control in healthy cats. A high-protein diet was associated with significantly elevated 41 cholesterol and triglyceride concentrations amongst healthy cats even though the increase was significantly 42 less pronounced in cats with a body condition score >5. This finding suggests that overweight cats process 43 high-protein diets, cholesterol and triglycerides differently than leaner cats.

- 44
- 45
- 46

48 **1.Introduction**

49

50 The cat is an obligate carnivore with various nutritional peculiarities adapted to a diet high in protein and 51 low in carbohydrates ¹¹. While older research postulated that high-carbohydrate diets increased the risk for 52 obesity in cats ²², more recent literature has presented contrasting findings ³³. Obesity in cats is associated 53 with an increased risk for the development of diabetes mellitus (DM)²². Dietary therapy for diabetic cats 54 focuses on reducing obesity, increasing muscle mass, decreasing postprandial hyperglycaemia and 55 controlling blood glucose fluctuations by minimising the need for β -cells to produce insulin⁴⁴. Current 56 evidence suggests that a high-protein, low-carbohydrate diet – relative to a high-carbohydrate diet – can 57 benefit cats with DM 5-8. Furthermore, clinical signs, blood glucose measurements and fructosamine 58 concentrations can be used to monitor glycaemic control and response to therapy ⁹⁻¹⁵. While researchers 59 agree that high-protein diets can help manage DM and obesity in cats ^{5-8, 16}, there is limited literature as to 60 whether this type of diet would be advantageous to the healthy cat.

61

62 There are conflicting reports about how carbohydrates and fats influence the glycaemic response in healthy cats. High-fat diets are associated with diminished glucose clearance and ß cell function ¹⁷ in contrast to 63 64 high-carbohydrate diets ^{18, 19}. There is also contradicting evidence regarding the influence of fibre on 65 glycaemic control in cats. While some have demonstrated better glycaemic control ²⁰, others have failed to 66 replicate these findings ⁶. Most of the research that has assessed how diet composition influences the feline 67 glycaemic response has focused on DM ^{5, 6, 8, 20}, with only a few studies using healthy cats ^{17, 18, 21, 22}. 68 Furthermore, these investigations differ widely in study design, feeding protocols, population size as well 69 as diet composition, which makes comparisons between studies difficult.

71	Several studies have reported that a high-protein and low-carbohydrate diet does not significantly affect
72	serum triglyceride concentrations in cats ^{19, 23, 24} . Comparatively, cats fed diets high in fat had significant
73	increases in triglyceride concentrations ^{17, 25, 26} . Although high-fibre foods lower triglyceride concentrations
74	in diabetic cats ²⁰ they increase cholesterol concentrations in healthy cats ²⁷ . Previous studies evaluating the
75	effects of high-fat diets on cholesterol concentrations have been contradictory. Some have reported that
76	high-fat diets do not contribute to hypercholesterolaemia ^{25, 28, 29} in comparison to others in which high fat
77	diets did contribute to hypercholesterolaemia in healthy cats ¹⁷ . Moreover, diabetic cats with increased
78	serum cholesterol concentrations are 65% less likely to achieve diabetic remission than cats with normal
79	serum cholesterol concentrations ³⁰ . This suggests that hypercholesterolaemia plays a primary role in the
80	progression of diabetes in cats by possibly preventing the recovery of β -cell function ³⁰ . This theory is
81	supported by several murine studies that have shown that elevated cholesterol concentrations can impair ß-
82	cell function ^{31, 32} . Current literature has also reported increased cholesterol concentrations in lean,
83	overweight and diabetic cats on the traditional high-protein diet prescribed to diabetic cats ²³ .
84	
85	The aim of this study was to determine the effect of three diets on serum cholesterol, triglyceride and
86	fructosamine concentrations in lean, normal and overweight non-diabetic cats.
87	
88	2. Materials and methods
89	
90	2.1. Experimental design
91	
92	The study was a randomised, crossover clinical trial that was approved by the Animal Ethics Committee of
93	the University of Pretoria (V079-18).

95	2.2. Animals
96	
97	Forty cats were recruited from an animal shelter, with thirty-five cats completing the study. Additionally,
98	of the forty cats, 30 were female and 10 were male and all were neutered. Equal numbers of cats were not
99	chosen, because animal selection depended on the availability of cats at the animal shelter. Three cats were
100	excluded due to early renal insufficiency while a further two were excluded due to behaviour-related issues.
101	To assist in proper identification, all cats were microchipped (Backhome, Virbac RSA., Centurion, South
102	Africa). The inclusion criteria were:
103	• Age over one year
104	• Not affected with overt renal or liver disease, DM, or hyperthyroidism
105	• Negative for feline immunodeficiency and leukaemia virus (FIV and FeLV)
106	• Not on any concurrent medical therapy
107	• Acceptance of restraint, venipuncture, and all diets
108	• No history of chronic vomiting or diarrhoea
109	• Known birthdate and complete vaccination record.
110	
111	2.3. Feeding Protocol
112	
113	All diets used are detailed in table 1. Prior to enrolment, participating cats were fed a commercial
114	maintenance/baseline diet by the animal shelter. Following baseline health assessments, cats were
115	randomised into groups that were fed either a high-protein or high-carbohydrate diet for four weeks. After
116	these four weeks, cats were fed a washout diet for four further weeks. The principal limitation of a crossover
117	trial is that the effects of one treatment may "carry over" and alter the response to subsequent treatments.
118	Thus, to prevent this, a washout diet which was an intermediate between the high-protein and high-

119 carbohydrate diet was chosen. Additionally, 4 weeks were selected to mimic the same amount of time that 120 the cats spent on each of the study diets. Thereafter, they were transitioned to the cross-over diet (Figure 121 1). Each cat was transitioned between diets over seven days. Cats were fed ad libitum of the dry and wet 122 diets. Body condition score (BCS) was determined based on the nine-point BCS chart ³³. Clinical 123 examinations, BCS, weight and environmental temperature (non-contact infrared thermometer, 124 Electromann SA, Pretoria, South Africa) measurements, were conducted on a weekly basis. Both dry and 125 wet high-protein diets were offered to all cats while only a dry diet for the high-carbohydrate and washout 126 diet.

127

Table 1. Comparison of the diets (reported on a dry matter basis) used in 35 healthy cats enrolled in a

129 cross-over study investigating the effects of diet on serum lipid and hormone profiles

Type of	Manufacturer	Protein	Fat	Carbohydrate	Crude
diet					Fibre
		21.10/	12.00/	12.00/	2.20/
Maintenan	Whiskas Beef,	31.1%	12.8%	43.9%	3.3%
ce	Lamb and				
	Rabbit flavour				
	with meaty				
	nuggets				
High-	Hill's M/D dry	50%	20.5%	19.6%	3.6%
protein	food				

High-	Hill's M/D wet	50.1%	21%	16.3%	7%
protein	food				
High-	Hill's Science	34.5%	10.3%	48.5%	1.1%
carbohydr	plan, Feline				
ate	mature adult 7+				
	sterilized cat				
Washout	Hill's Science	34.3%	20%	31.5%	8.6%
	Plan, Feline				
	Mature Adult				
	7+ Hairball				
	Control)				



150 BCS = body condition score; CBC=complete blood count; TT4=total thyroxine.

151 Figure 1: Flow chart highlighting the research process in 40 healthy cats randomised into groups receiving

152 either a high-carbohydrate or high-protein diet first during a cross-over study investigating the effects of

153 diet on serum lipid and hormone profiles

154

155 2.4. Health Assessment and Laboratory Tests

156

157 Cats were determined as healthy based on history, physical examination, and laboratory tests. Blood 158 samples were collected from all forty cats prior to the start of the study after a 12 hour fast. Blood was 159 collected from the jugular vein by needle venipuncture and placed into one serum and one EDTA tube. 160 Serum cholesterol, triglyceride, alkaline phosphatase (ALP), alanine aminotransferase (ALT), gamma-161 glutamyl transferase (GGT), blood urea nitrogen (BUN), creatinine, glucose, albumin, globulin and total 162 serum protein (TSP) levels were measured using the Cobas Integra 400 plus analyser (Roche Diagnostics, 163 Risch-Rotkreuz, Switzerland). The ADVIA 2120 Hematology System (Siemens Healthineers, Erlangen, 164 Germany) was used to obtain all complete blood counts (CBCs). Total thyroxine (TT4) concentrations were 165 measured with the Immulite 2000 immunoassay system (Siemens Healthineers). Fructosamine 166 concentration was determined using a colorimetric method on the Cobas Integra 400 plus analyser. A SNAP 167 Combo plus (Idexx Laboratories), enzyme-linked immunosorbent assay for the simultaneous detection of 168 FeLV antigen and antibodies for FIV was performed. All biochemistry and complete blood counts were 169 performed at the Clinical Pathology Laboratory at the Faculty of Veterinary Science, University of Pretoria. 170 Fructosamine concentrations were determined at a commercial laboratory (Idexx Laboratories, 171 Johannesburg, South Africa). Blood samples were collected from the cats after the four-week feeding period 172 with the high-protein, high-carbohydrate and washout diets. Blood samples were centrifuged and serum 173 centrifuged, separated and refrigerated within 1 hour of collection. Thereafter, serum was aliquoted and 174 frozen at 80°C within 24 hours of collection. At the end of the study, all collected specimens were analysed

in a single batch.

176

177 2.6. Statistical Analyses

178

179 Data were assessed for normality of distribution by plotting histograms, calculating descriptive statistics 180 and performing the Anderson-Darling test (MINITAB Statistical Software, Release 13.32, Minitab Inc, 181 State College, Pennsylvania, USA). Right-skewed data were transformed using the natural logarithm. 182 Categorical data were described using proportions and 95% confidence intervals (CI) while quantitative 183 data were described using medians and interquartile ranges (IQR). Quantitative data were further evaluated 184 by creating boxplots using the ggplot2 package (Wickham, 2009) within R (R Development Core Team, 185 2017). Categorical data were compared between cats based on the first diet assignment groups using chi-186 square tests (Epi Info, version 6.04, CDC, Atlanta, GA). Quantitative data were compared between initial 187 diet assignment groups using independent t-tests on the raw or natural logarithm transformed data. Mann-188 Whitney U tests were used when the normality assumption was violated. Mixed-effects linear models were 189 created to determine the effect of diet and BCS on serum cholesterol, triglyceride and fructosamine 190 concentrations. Cat was included as a random effect in all models and the correlation among repeated 191 measures was modelled using a first-order autoregressive (AR1) covariance structure. Evaluated fixed 192 effects included diet, ordinal BCS groupings, sex, breed, age, experimental room, room temperature, and 193 pairwise interactions between BCS and diet. Complete models were fit and a backwards stepwise process 194 was employed to remove predictors with the largest P values until all remaining variables had significant 195 slope parameters. Unless otherwise stated, SPSS (IBM SPSS Statistics Version 25, International Business 196 Machines Corp., Armonk, NY, USA) was used for all statistical analyses. Significance was set at p < 0.05. 197

198	3. Results
199	
200	3.1. Baseline Data
201	
202	There were no statistical differences in the baseline data between the two initial diet groups (Table 2 and
203	3).
204	
205	Table 2. Comparison of baseline demographics and serum chemistry parameters in 40 healthy cats
206	randomised into groups receiving either a high-carbohydrate (HC; n=20) or high-protein (HP; n=20) diet
207	first during a cross-over study investigating the effects of diet on serum lipid and hormone profiles

	HC diet first		HP diet first			
Variable	n/d	PE* (Interval†)	n/d	PE* (Interval†	P value‡	
)		
Categorical data						
Female sex	15/20	0.75 (0.53, 0.90)	15/20	0.75 (0.53, 0.90)	1.0‡	
DSH	17/20	0.85 (0.64, 0.96)	12/20	0.60 (0.38, 0.79)	0.07‡	
Quantitative data						
Age (yr)	20/20	4 (2,7)	20/20	4 (3, 6)	0.84§	
Albumin (g/L)	20/20	33.1 (32.3, 34.4)	20/20	34.0 (31.2, 37.7)	0.80¶	
ALP (U/L)	20/20	35.5 (26.8, 44.8)	20/20	31.5 (24.3, 45.5)	0.87¶	
ALT (U/L)	20/20	43. 7 (35.2, 54.2)	20/20	40.3 (33.8, 49.4)	0.31¶	
BCS (/9)	20/20	5 (4, 6)	20/20	5 (4, 6)	1.0¶	

BUN (mmol/L)	20/20	7.1 (5.9, 9.0)	20/20	7.8 (6.2, 9.8)	0.34¶
Cholesterol (mmol/L)	20/20	2.46 (1.98, 3.02)	20/20	2.46 (1.99, 2.76)	0.80#
Creatinine (umol/L)	20/20	112 (104, 121)	20/20	118 (104, 131)	0.39¶
Fructosamine (mmol/L)	20/20	247 (227, 270)	20/20	239 (214, 275)	0.44#
GGT (U/L)	20/20	0 (0, 1)	20/20	0 (0, 0)	0.12§
Globulin (g/L)	20/20	39.7 (34.4, 43.4)	20/20	36.8 (33.8, 41.7)	0.30¶
Glucose (mmol/L)	20/20	4.2 (3.8, 4.6)	20/20	4.1 (3.7, 4.6)	0.86§
Triglycerides (mmol/L)	20/20	0.39 (0.31, 0.52)	20/20	0.33 (0.27, 0.38)	0.17#
TSP (g/L)	20/20	73.5 (69.9, 77.1)	20/20	70.2 (67.5, 74.6)	0.36¶
TT4 (nmol/L)	20/20	22.1 (20.1, 26.4)	20/20	23.6 (21.1, 29.4)	0.93#
Weight (kg)	20/20	3.7 (3.6, 4.3)	20/20	3.9 (3.5, 4.2)	0.99#

209 n/d = numerator/denominator; ALP = alkaline phosphatase; ALT = alanine transaminase; BCS = body

210 condition score; BUN = blood urea nitrogen; CI = confidence interval; DSH = domestic short hair breed;

211 GGT = gamma-glutamyl transferase; TSP = total serum protein; TT4 = Total thyroxine.

212 *PE = point estimate, corresponding to the proportion for categorical variables and the median for

- 213 quantitative data
- ²¹⁴ †Interval is the 95% confidence interval for categorical data and the interquartile range for quantitative data
- 215 ‡Based on chi-square tests
- 216 §Based on Mann-Whitney U tests
- 217 Based on independent t-tests on untransformed data
- 218 #Based on independent t-test on natural log-transformed data

220 **Table 3.** Comparison of complete blood count results in 40 healthy cats randomised into groups receiving

221 either a high-carbohydrate (HC; n=20) or high-protein (HP; n=20) diet first during a cross-over study

- 222 investigating the effects of diet on serum lipid and hormone profiles
- 223

	HC diet first		HP diet	HP diet first	
Variable	n/d	Median (IQR)	n/d	Median (IQR)	P value
Band neutrophils (×10 ⁹ /L)	16/20	0 (0, 0.05)	17/20	0 (0, 0)	0.81*
Basophils (×10 ⁹ /L)	16/20	0 (0, 0)	17/20	0 (0, 0)	0.76*
Eosinophils (×10 ⁹ /L)	16/20	0.44 (0.26, 0.88)	17/20	0.54 (0.42, 0.76)	0.91†
Hematocrit (L/L)	16/20	0.35 (0.31, 0.38)	17/20	0.37 (0.32, 0.40)	0.59‡
Hemoglobin (g/L)	16/20	125 (121, 139)	17/20	140 (113, 147)	0.60‡
Lymphocytes (×10 ⁹ /L)	16/20	3.61 (3.10, 5.07)	17/20	3.39 (2.76, 5.41)	0.68*
MCHC (g/dL)	16/20	36.8 (35.5, 38.7)	17/20	36.5 (35.6, 37.5)	0.73‡
MCH (pg)	16/20	15.7 (14.4, 16.2)	17/20	14.1 (13.4, 15.8)	0.17†
MCV (fL)	16/20	41.6 (38.8, 44.2)	17/20	39.2 (37.3, 42.3)	0.24‡
Monocytes (×10 ⁹ /L)	16/20	0.29 (0.16, 0.60)	17/20	0.26 (0.11, 0.39)	0.38†
Neutrophils (×10 ⁹ /L)	16/20	5.97 (4.29, 9.25)	17/20	6.25 (5.51, 7.95)	0.71†
Platelets (×10 ⁹ /L)	16/20	204 (151, 316)	17/20	316 (189, 523)	0.10†
RCC (×10 ¹² /L)	16/20	8.49 (7.52, 9.81)	17/20	8.81 (7.61, 10.60)	0.38*
RDW %	16/20	14.5 (14.0, 15.2)	17/20	14.7 (14.4, 15.1)	0.83‡
WCC (×10 ⁹ /L)	16/20	11.9 (8.2, 13.6)	17/20	10.6 (9.0, 14.0)	0.86†

224 n/d = numerator / denominator; IQR = interquartile range; MCH= mean corpuscular haemoglobin; MCHC=

225 mean corpuscular haemoglobin concentration; MCV= mean corpuscular volume; RCC= red cell count;

226 RDW= red cell distribution width; WCC= white cell count.

227	*Based on Mann-Whitney U tests
228	†Based on independent t-test on natural log-transformed data
229	‡Based on independent t-tests on untransformed data
230	
231	3.2. Body Weight
232	
233	Having been fed a high-protein diet (P=0.001), being male (P<0.001) and having a BCS>5 (P=0.002) were
234	significant predictors of heavier body weights (Table 4).
235	
236	Table 4. Multivariable associations between body weight*, diet, and body condition score (BCS) in 35
237	healthy cats enrolled in a cross-over study investigating the effects of diet on serum lipid and hormone
238	profiles
239	

Variable	Level	Estimate (95% CI)	t statistic	P value
Diet	НС	0.005 (-0.016, 0.026)	0.477	0.63
	HP	0.036 (0.015, 0.058)	3.421	0.001
	Washout	0.025 (-0.001, 0.050)	1.920	0.05
	Baseline	Reference		
Sex	Male	0.187 (0.103, 0.270)	4.531	< 0.001
	Female	Reference		
BCS < 5	Yes	-0.071 (-0.104, -0.038)	-4.250	< 0.001
	No	Reference		

BCS > 5	Yes	0.047 (0.017, 0.078)	3.100	0.002
	No	Reference		
Room temperature	1 C increase	-0.009 (-0.015, -0.003)	-2.812	0.006

240 $\overline{\text{CI} = \text{confidence interval; HC} = \text{high carbohydrate; HP} = \text{high protein.}}$

241 *Data were natural log-transformed prior to statistical analysis

242 **3.3. Cholesterol**

243

244 Median cholesterol concentrations were highest on the high-protein diet (Figure 2; Table 5). Cholesterol 245 concentrations were above the reference range in 10% (15/145) of the samples, of which 87% (13/15) 246 represented the high-protein diet and 13% (2/15) the washout diet. The cholesterol concentrations of the 247 remainder of the samples, 90% (130/145), were either within or just below the reference range. Cats on the 248 high-carbohydrate diet had significantly lower (P<0.001) cholesterol concentrations than cats on either the 249 high-protein (P<0.001) or washout diets (P<0.001; Table 6). Moreover, cats with a BCS >5 and that were 250 fed a high-protein diet had significantly lower (P=0.007) cholesterol concentrations than cats from other 251 BCS groups.

252



254 Figure 2. Illustration of serum cholesterol values for 35 healthy cats – separated according to body

255 condition score (BCS) - in a cross-over study investigating the effects of diet on serum lipid and hormone

256 profiles. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends

of boxes), and 10th and 90th percentiles (perpendicular lines)

258

259	Table 5. Descriptive statistics for 35 healthy cats in an experimental cross-over study investigating the
260	effects of diet on serum lipid and hormone profiles.
261	

	Cholest	Cholesterol (mmol/L)		Fructosamine (mmol/L)		Triglyceride (mmol/L)	
Variable	n	Median (IQR)	n	Median (IQR)	n	Median (IQR)	
Lean cats (BCS <5)							
Baseline	13	2.46 (1.87, 2.78)	13	256 (228, 278)	13	0.35 (0.27, 0.48)	
HC diet	12	1.97 (1.57, 2.43)	12	229 (215, 242)	12	0.31 (0.26, 0.43)	
HP diet	10	3.91 (2.94, 4.45)	10	209 (198, 258)	10	0.36 (0.29, 0.59)	
Washout diet	10	2.95 (1.85, 3.14)	10	206 (201, 230)	10	0.28 (0.23, 0.34)	
Normal cats (BCS = 5)							
Baseline	9	2.56 (2.17, 3.10)	9	241 (216, 267)	9	0.32 (0.29, 0.47)	
HC diet	13	2.26 (1.81, 2.42)	13	238 (207, 271)	13	0.40 (0.31, 0.58)	
HP diet	11	3.58 (2.95, 5.10)	11	220 (204, 232)	11	0.45 (0.35, 0.59)	
Washout diet	14	2.91 (2.09, 3.61)	12	204 (194, 224)	14	0.29 (0.24, 0.33)	
Overweight cats (BCS > 5)							
Baseline	13	2.45 (2.16, 2.67)	13	255 (201, 278)	13	0.38 (0.29, 0.47)	
HC diet	10	2.43 (1.90, 2.67)	10	241 (200, 260)	10	0.40 (0.37, 0.51)	
HP diet	14	4.05 (2.79, 4.45)	14	214 (201, 235)	14	0.39 (0.37, 0.50)	
Washout diet	11	3.42 (2.41, 3.78)	11	220 (203, 232)	11	0.33 (0.31, 0.38)	

 $26\overline{2}$ IQR = interquartile range. HC = high carbohydrate. HP = high protein.

263

264

Table 6. Multivariable associations between serum cholesterol*, diet, and body condition score (BCS) in
35 healthy cats enrolled in a cross-over study investigating the effects of diet on serum lipid and hormone
profiles

Variable	Level	Estimate (95% CI)	t statistic	P value
Diet	HC	-0.122 (-0.186, -0.059)	20.628	< 0.001
	HP	0.479 (0.408, 0.551)	-3.814	< 0.001
	Washout	0.166 (0.099, 0.233)	13.323	< 0.001
	Baseline	Reference		
BCS > 5	Yes	0.001 (-0.085, 0.088)	0.031	0.97
	No	Reference		
BCS > 5 and HP diet	Yes	-0.157 (-0.269, -0.044)	-2.766	0.007
	No	Reference		

270 CI = confidence interval; HC = high carbohydrate; HP = high protein.

271 *Data were natural log-transformed prior to statistical analysis

272

269

273

274 **3.4. Triglycerides**

275

276 Median triglyceride concentrations were lowest on the washout diet (Figure 3; Table 5). None of the 277 triglyceride concentrations were above the reference range. Cats that had been fed the washout diet had 278 significantly lower (P=0.009) concentrations of triglycerides, whereas cats fed the high-protein diet had

- significantly higher (P<0.001; Table 7) concentrations. Cats with a BCS >5 and fed a high-protein diet had
- significantly (P=0.03) lower triglyceride concentrations than those from other BCS groups.
- 281



282

Figure 3. Illustration of serum triglyceride values for 35 healthy cats – separated according to body condition score (BCS) - in a cross-over study investigating the effects of diet on serum lipid and hormone profiles. Data are shown as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and 90th percentiles (perpendicular lines)

Table 7. Multivariable associations between serum triglycerides*, diet, and body condition score (BCS)
in 35 healthy cats enrolled in a cross-over study investigating the effects of diet on serum lipid and hormone

- 290 profiles
- 291

Variable	Level	Estimate (95% CI)	t statistic	P value
Diet	HC	0.068 (-0.029, 0.164)	1.386	0.16
	HP	0.223 (0.114, 0.332)	4.045	< 0.001
	Washout	-0.134 (-0.233, -0.034)	-2.669	0.009
	Baseline	Reference		
BCS > 5	Yes	-0.009 (-0.131, 0.112)	-0.152	0.87
	No	Reference		
BCS > 5 and HP diet	Yes	-0.192 (-0.366, -0.017)	-2.180	0.03
	No	Reference		

292 CI = confidence interval; HC = high carbohydrate; HP = high protein.

293 *Data were natural log-transformed prior to statistical analysis

294

3.5. Fructosamine

296

Median fructosamine concentrations were highest in cats fed the high-carbohydrate diet (Figure 4; Table
5). Only 6% (9/143) of the fructosamine samples were below the reference range; of these, four represented
cats on the high-protein diet, four on the washout diet, and one on the baseline diet. The remaining 94%
(134/143) of samples were within the reference range. Cats on either the high-protein or washout diets had
significantly lower (P<0.001) fructosamine concentrations than other cats (Table 8).



150 -



Figure 4. Illustration of serum fructosamine values for 35 healthy cats – separated by body condition score
- in a cross-over study investigating the effects of diet on serum lipid and hormone profiles. Data are shown
as median (horizontal line within box), 25th and 75th percentiles (horizontal ends of boxes), and 10th and
90th percentiles (perpendicular lines)

309 Table 8. Associations between serum fructosamine* and diet in 35 healthy cats enrolled in a cross-over

310	study investigating the effects of	diet on serum lipid and hormone profiles

HC	-0.037 (-0.077, 0.002)	-1.878	0.06
ΗP	-0.110 (-0.149, -0.070)	-5.514	< 0.001
Washout	-0.133 (-0.169, -0.097)	-7.338	< 0.001
Baseline	Reference		
	HC HP Washout Baseline	HC -0.037 (-0.077, 0.002) HP -0.110 (-0.149, -0.070) Washout -0.133 (-0.169, -0.097) Baseline Reference	HC -0.037 (-0.077, 0.002) -1.878 HP -0.110 (-0.149, -0.070) -5.514 Washout -0.133 (-0.169, -0.097) -7.338 Baseline Reference

311 CI = confidence interval; HC = high carbohydrate; HP = high protein.

312 *Data were natural log-transformed prior to statistical analysis. No multivariable model fit the data.

313

314 **4. Discussion**

315

This study showed that cats on a high-carbohydrate diet had significantly lower serum cholesterol concentrations than cats on the maintenance diet. Cats on a high-protein diet had significantly higher serum cholesterol and triglyceride concentrations, yet lower fructosamine concentrations compared to baseline measurements. In contrast, overweight cats (BCS>5) had lower cholesterol and triglyceride concentrations on a high-protein diet than cats representing other BCS groups.

321

322 It has been shown that neutered male cats are at an increased risk of obesity compared to intact males, and 323 therefore at greater risk for developing DM ^{22, 34, 35}. In the current study, neutered male cats were 324 significantly heavier than neutered female cats, which is in agreement with previous findings ^{36, 37}. It has 325 been postulated that high-carbohydrate diets increase the risk for obesity in cats ²²; however, there are few 326 epidemiological studies available that either support or refute this claim ³⁵. In the current study, cats on a 327 high-carbohydrate diet were not heavier than cats on other diets. This supports prior reports that cats limit 328 their total energy intake when consuming a high-carbohydrate diet ^{18, 38}. However, lack of weight gain in 329 this study could of resulted from a lack of uniformity in caloric intake between groups and the time on 330 each diet. Dietary protein is an important component of weight loss diets ³⁹, as high-protein diets have been 331 shown to promote fat loss in cats ¹⁶. However, offering overweight cats an *ad libitum* high-protein diet 332 increases food intake – perhaps due to increased palatability – without any noticeable changes in body weight or composition ²⁴. Our *ad libitum* experiment demonstrated that cats fed a high-protein diet were 333 334 heavier than cats on other diets.

335 In DM cats, hypercholesterolaemia reduces the chance of remission by almost 65%. Although 336 hypercholesterolaemia can contribute to the pathogenesis of DM in cats ³⁰, its effects on healthy cats are 337 still debateable. Prior research has reported that diabetic, lean and overweight cats fed a high-protein diet 338 had higher serum cholesterol concentrations²³. In this current study, cats that were fed a high-protein diet 339 had significantly higher cholesterol concentrations compared to other cats. Cats fed the high-protein diet 340 had elevated median cholesterol concentrations among all three BCS groups. Interestingly, overweight cats 341 on the high-protein diet did not have a large increase in serum cholesterol concentrations. The mechanism 342 through which ingested protein is coupled to upregulated cholesterol production requires further study.

343

344 Studies have indicated that insoluble fibre is positively associated with cholesterol concentrations in 345 overweight cats. It has been speculated that fibre can interfere with the absorption of specific fat 346 components that could subsequently alter which lipoproteins are synthesised in the liver ²⁷. It should be 347 noted that the high-protein diet of the current study had considerably higher crude insoluble fibre content 348 than the high-carbohydrate diet. The high-protein diet had nearly double the amount of fat that was in the 349 high-carbohydrate diet. It has been reported that a high-fat diet does not contribute to hypercholesterolaemia 350 ^{25, 28, 29}. Nevertheless, prior studies of cats fed a high-fat diet reported higher cholesterol concentrations 351 compared to cats fed a high-carbohydrate diet ¹⁷. Cats in the current study fed the high-carbohydrate diet, 352 which has low fibre, protein and fat content, had lower cholesterol concentrations while cats fed the high-353 protein diet, which has high fibre, protein and fat content, had increased cholesterol concentrations. As 354 interactions between different dietary components may have exerted an additive role in these findings; 355 further studies are required to specifically address lipoprotein fractions.

357 There are conflicting reports of the influence of diet on triglyceride concentrations in cats. Several studies 358 reported that a high-protein and low-carbohydrate diet did not significantly affect triglyceride 359 concentrations in cats ^{19, 23, 24}; whereas others have shown that a high-fat diet had a significant increase in 360 triglyceride concentrations ^{17, 25, 26}. This current study showed that a high-protein diet increased triglyceride 361 concentrations in healthy cats. The discrepancies between these findings and previous reports might be due 362 to the differences in study design. Overweight cats generally have higher triglyceride concentrations than 363 healthy cats ⁴⁰. In this current study, lean and normal cats fed a high-protein diet and overweight cats fed a 364 high-carbohydrate diet had the highest median triglyceride concentrations. In this current study, cholesterol 365 and triglyceride concentrations in overweight cats decreased when fed a high-protein diet.

366

367 Diabetic cats can benefit from high-protein and low-carbohydrate diets due to higher diabetic remission 368 rates ⁶, lower fructosamine concentrations ⁸, and improved glycaemic control ^{5, 8}. The majority of studies 369 on the effect of diet composition on feline glycaemic response have focused on diabetic cats ^{5-8, 20}, with 370 only a few studies involving healthy cats ^{17, 18, 21, 22}. This current study showed that cats on the high-371 carbohydrate diet tended to have the highest median fructosamine concentrations. In the literature, there 372 are conflicting reports on the effect of carbohydrates and fats on the glycaemic response in healthy cats. 373 One study showed that cats fed a high-fat diet had a diminished glucose clearance and ß-cell function 374 relative to cats fed a high-carbohydrate diet ¹⁷. There is also evidence that high-carbohydrate diets cause 375 higher insulin ²⁶ and post-prandial glucose concentrations^{18, 19} compared to healthy cats fed high-fat and 376 high-protein diets. There seems to be a complex link between diet and fructosamine concentrations among 377 healthy cats; more specifically, this current study revealed that carbohydrate and fat contents were 378 positively and negatively, respectively, linked with fructosamine concentrations in healthy cats.

380 In this current study, cats on both high-protein and washout diets had significantly lower fructosamine 381 concentrations compared to cats on the other diets. These findings agree with the finding that healthy cats 382 fed high-protein diets with either low or moderate levels of starch had significantly decreased glucose and 383 fructosamine concentrations in comparison to moderate-protein and high-starch diets⁴¹. There is conflicting 384 evidence regarding the influence of fibre on the glycaemic control in cats. Glycaemic control among DM 385 cats improved when fibre intake was increased to moderate levels 20 in contrast to other studies that showed 386 the opposite ⁶. Cats continue with post-prandial gluconeogenesis from protein, which might explain why 387 fibre is potentially less effective in this species ^{42, 43}. Even though the exact mechanisms underlying these 388 findings remain unknown, it is speculated that dietary fibre affects the nutrient transit rate in the gut, which, 389 subsequently, reduces glucose absorption along with post-prandial glycaemia and enhances glycaemic 390 control ^{20, 44}. This current study suggests that diets high in protein, fibre, and fat, but low in carbohydrates, 391 could contribute to decreased glucose concentrations in healthy cats.

392

393 This study had several limitations. The included cats were not fed according to their individual nutritional 394 requirements, but ad libitum to simulate the situation in private, multi-cat households. Ad libitum feeding 395 is regarded as a risk factor for obesity, and thus the feeding strategy in this study might have inadvertently 396 predisposed the participating cats to gain weight ⁴⁵. The amount of food that each cat ingested was not 397 recorded and thus some cats might have preferred one type of food to another, which could have introduced 398 bias. Additionally, a hierarchical structure will occur with group housing of cats with dominant animals 399 eating more and submissive animals eating less. While this might have introduced bias, an adaptation period 400 of 2 months to identify these cats was performed to reduce this limitation. Two such cats were identified 401 and excluded. Finally, wet food was only used for the high-protein diet and this could have influenced 402 presented findings; however, the dietary contents of the wet and dry high-protein diets had very similar 403 protein, carbohydrate and fat composition. Additionally, both types of feed(wet and dry) of this specific

404	diet would traditionally be given to a cat in the clinical setting, and we attempted to replicate the decision
405	a clinician would face in private practice.
406	
407	5. Conclusions:
408	
409	In conclusion, diets with high protein, but low carbohydrate content, might be beneficial for short-term
410	glucose control in healthy cats. The reduction in cholesterol and triglyceride concentrations among
411	overweight cats on a high-protein diet, relative to lean and normal cats on the same diet, is a novel result
412	that warrants further investigation. The finding that a high-protein diet significantly increased cholesterol
413	and triglyceride concentrations and a high-carbohydrate diet significantly decreased cholesterol
414	concentrations in healthy cats relative to other diets also warrants further investigation.
415	
416	Acknowledgements
417	
418	The authors thank the staff of the Animal-Anti Cruelty League for the care of the animals and usage of their
419	facilities and nurses Jade Little and Roxanne Lundin for their assistance.
420	
421	Author notes
422	Presented as a poster presentation at the annual ECVIM-CA online congress, 2-5 September 2020.
423	
424	Conflict of Interest

427	publication of this article.
428	
429	Funding
430	
431	This work was supported by the Pathobiology research theme of the Faculty of Veterinary Science of the
432	University of Pretoria, the South African Veterinary Foundation [grant number SAVF 1652]; the Health
433	and Welfare Sector Education and Training Authority [grant number 2808743]. Hills provided all the
434	food and Virbac provided all the microchips. None of the funders and companies mentioned above had
435	any role in the design, analysis and writing of this article.
436	
437	Ethical Approval
438	
439	This work involved the use of experimental animals and the study therefore had ethical approval from an
440	established committee as stated in the manuscript.
441	
442	Informed consent
443	Informed consent (either verbal or written) was obtained from the owner or legal custodian of all
444	animal(s) described in this work (either experimental or non-experimental animals) for the procedure(s)
445	undertaken (either prospective or retrospective studies). No animals or humans are identifiable within this
446	publication, and therefore additional informed consent for publication was not required.
447	
448	References

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or

450 1. Morris JG. Idiosyncratic nutrient requirements of cats appear to be diet-induced

451 evolutionary adaptations. *Nutr Res Rev* 2002; 15: 153-168. 2002/06/01. DOI: 10.1079/nrr200238.

- 452 2. Scarlett JM, Donoghue S, Saidla J, et al. **Overweight cats: prevalence and risk factors**.
- 453 International journal of obesity and related metabolic disorders : journal of the International Association
- 454 *for the Study of Obesity* 1994; 18 Suppl 1: S22-28. 1994/06/01.
- 455 3. Cave NJ, Allan FJ, Schokkenbroek SL, et al. A cross-sectional study to compare changes in

456 the prevalence and risk factors for feline obesity between 1993 and 2007 in New Zealand. *Preventive*

- 457 *veterinary medicine* 2012; 107: 121-133. 2012/06/19. DOI: 10.1016/j.prevetmed.2012.05.006.
- 458 4. Zoran DL and Rand JS. The role of diet in the prevention and management of feline
- 459 diabetes. The Veterinary clinics of North America Small animal practice 2013; 43: 233-243. 2013/03/26.
- 460 DOI: 10.1016/j.cvsm.2012.11.004.
- 461 5. Mazzaferro EM, Greco DS, Turner AS, et al. Treatment of feline diabetes mellitus using an

462 **alpha-glucosidase inhibitor and a low-carbohydrate diet**. *Journal of feline medicine and surgery* 2003;

463 5: 183-189. 2003/05/27. DOI: 10.1016/s1098-612x(03)00006-8.

464 6. Bennett N, Greco DS, Peterson ME, et al. Comparison of a low carbohydrate-low fiber diet

465 and a moderate carbohydrate-high fiber diet in the management of feline diabetes mellitus. *Journal*

466 *of feline medicine and surgery* 2006; 8: 73-84. 2005/11/09. DOI: 10.1016/j.jfms.2005.08.004.

- 467 7. Frank G, Anderson W, Pazak H, et al. Use of a high-protein diet in the management of feline
- 468 diabetes mellitus. Veterinary therapeutics : research in applied veterinary medicine 2001; 2: 238-246.
 469 2001/07/01.
- 470 8. Hall TD, Mahony O, Rozanski EA, et al. Effects of diet on glucose control in cats with
- 471 **diabetes mellitus treated with twice daily insulin glargine**. *Journal of feline medicine and surgery*
- 472 2009; 11: 125-130. 2008/10/07. DOI: 10.1016/j.jfms.2008.06.009.

473	9.	Crenshaw KL	, Peterson ME, He	eb LA, et al.	. Serum fruct	osamine conce	ntration as	an inde	X
-----	----	-------------	-------------------	---------------	---------------	---------------	-------------	---------	---

- 474 of glycemia in cats with diabetes mellitus and stress hyperglycemia. Journal of veterinary internal
- 475 *medicine* 1996; 10: 360-364. 1996/11/01. DOI: 10.1111/j.1939-1676.1996.tb02081.x.
- 476 10. Link KR and Rand JS. Changes in blood glucose concentration are associated with relatively
- 477 rapid changes in circulating fructosamine concentrations in cats. Journal of feline medicine and
- 478 *surgery* 2008; 10: 583-592. 2008/11/08. DOI: 10.1016/j.jfms.2008.08.005.
- 479 11. Plier ML, Grindem CB, MacWilliams PS, et al. Serum fructosamine concentration in
- 480 nondiabetic and diabetic cats. *Veterinary clinical pathology* 1998; 27: 34-39. 2002/06/21. DOI:
- 481 10.1111/j.1939-165x.1998.tb01013.x.
- 482 12. Kaneko JJ, Kawamoto M, Heusner AA, et al. Evaluation of serum fructosamine
- 483 **concentration as an index of blood glucose control in cats with diabetes mellitus**. *American journal of*
- 484 *veterinary research* 1992; 53: 1797-1801. 1992/10/01.
- 485 13. Reusch CE, Liehs MR, Hoyer M, et al. Fructosamine. A new parameter for diagnosis and
- 486 **metabolic control in diabetic dogs and cats**. *Journal of veterinary internal medicine* 1993; 7: 177-182.
- 487 1993/05/01. DOI: 10.1111/j.1939-1676.1993.tb03183.x.
- 488 14. Lutz TA, Rand JS and Ryan E. Fructosamine concentrations in hyperglycemic cats. *The*
- 489 *Canadian veterinary journal = La revue veterinaire canadienne 1995; 36: 155-159. 1995/03/01.*
- 490 15. Martin GJ and Rand JS. Comparisons of different measurements for monitoring diabetic
- 491 cats treated with porcine insulin zinc suspension. *Vet Rec* 2007; 161: 52-58. 2007/07/17. DOI:
- 492 10.1136/vr.161.2.52.
- 493 16. P. Laflamme D and Hannah S. Increased Dietary Protein Promotes Fat Loss and Reduces Loss
- 494 of Lean Body Mass During Weight Loss in Cats. 2005.

495 17. Thiess S, Becskei C, Tomsa K, et al. Effects of high carbohydrate and high fat diet on

496 plasma metabolite levels and on i.v. glucose tolerance test in intact and neutered male cats. J Feline

497 *Med Surg* 2004; 6: 207-218. 2004/07/22. DOI: 10.1016/j.jfms.2003.09.006.

- 498 18. Farrow HA, Rand JS, Morton JM, et al. Effect of dietary carbohydrate, fat, and protein on
- 499 **postprandial glycemia and energy intake in cats**. *Journal of veterinary internal medicine* 2013; 27:
- 500 1121-1135. 2013/07/23. DOI: 10.1111/jvim.12139.
- 501 19. Mimura K, Mori A, Lee P, et al. Impact of commercially available diabetic prescription diets
- 502 on short-term postprandial serum glucose, insulin, triglyceride and free fatty acid concentrations of
- **503 obese cats**. *J Vet Med Sci* 2013; 75: 929-937. 2013/03/02.
- 504 20. Nelson RW, Scott-Moncrieff JC, Feldman EC, et al. Effect of dietary insoluble fiber on
- 505 control of glycemia in cats with naturally acquired diabetes mellitus. Journal of the American
- 506 *Veterinary Medical Association* 2000; 216: 1082-1088. 2000/04/08.
- 507 21. Mori A, Sako T, Lee P, et al. Comparison of three commercially available prescription diet
- 508 regimens on short-term post-prandial serum glucose and insulin concentrations in healthy cats.
- 509 *Veterinary research communications* 2009; 33: 669-680. 2009/03/27. DOI: 10.1007/s11259-009-9216-5.
- 510 22. Hoenig M, Thomaseth K, Waldron M, et al. Insulin sensitivity, fat distribution, and
- 511 adipocytokine response to different diets in lean and obese cats before and after weight loss.
- 512 American journal of physiology Regulatory, integrative and comparative physiology 2007; 292: R227-
- 513 234. 2006/08/12. DOI: 10.1152/ajpregu.00313.2006.
- 514 23. Zapata RC, Meachem MD, Cardoso NC, et al. Differential circulating concentrations of
- 515 adipokines, glucagon and adropin in a clinical population of lean, overweight and diabetic cats.
- 516 BMC Vet Res 2017; 13: 85. 2017/04/06. DOI: 10.1186/s12917-017-1011-x.

517 24. Wei A, Fascetti AJ, Liu KJ, et al. Influence of a high-protein diet on energy balance in obese

518 cats allowed ad libitum access to food. J Anim Physiol Anim Nutr (Berl) 2011; 95: 359-367.

519 2010/11/03. DOI: 10.1111/j.1439-0396.2010.01062.x.

520 25. Trevizan L, de Mello Kessler A, Bigley KE, et al. Effects of dietary medium-chain

521 triglycerides on plasma lipids and lipoprotein distribution and food aversion in cats. American

- 522 *journal of veterinary research* 2010; 71: 435-440. 2010/04/07. DOI: 10.2460/ajvr.71.4.435.
- 523 26. Keller C, Liesegang A, Frey D, et al. Metabolic response to three different diets in lean cats
- and cats predisposed to overweight. BMC Veterinary Research 2017; 13: 184. DOI: 10.1186/s12917-
- 525 017-1107-3.
- 526 27. Fischer MM, Kessler AM, de Sa LR, et al. Fiber fermentability effects on energy and

527 macronutrient digestibility, fecal traits, postprandial metabolite responses, and colon histology of

- 528 overweight cats. Journal of animal science 2012; 90: 2233-2245. 2012/01/17. DOI: 10.2527/jas.2011-
- 529 4334.
- 530 28. Butterwick RF, Salt C and Watson TD. Effects of increases in dietary fat intake on plasma

531 lipid and lipoprotein cholesterol concentrations and associated enzyme activities in cats. American

532 *journal of veterinary research* 2012; 73: 62-67. 2011/12/30. DOI: 10.2460/ajvr.73.1.62.

533 29. Jordan E, Kley S, Le NA, et al. Dyslipidemia in obese cats. *Domestic animal endocrinology*

534 2008; 35: 290-299. 2008/08/12. DOI: 10.1016/j.domaniend.2008.05.008.

535 30. Zini E, Hafner M, Osto M, et al. Predictors of clinical remission in cats with diabetes

536 mellitus. Journal of veterinary internal medicine 2010; 24: 1314-1321. 2010/09/16. DOI:

- 537 10.1111/j.1939-1676.2010.0598.x.
- 538 31. Hao M, Head WS, Gunawardana SC, et al. Direct effect of cholesterol on insulin secretion: a
- **539** novel mechanism for pancreatic beta-cell dysfunction. *Diabetes* 2007; 56: 2328-2338. 2007/06/19.
- 540 DOI: 10.2337/db07-0056.

541 32. Ishikawa M, Iwasaki Y, Yatoh S, et al. Cholesterol accumulation and diabetes in pancreatic

542 beta-cell-specific SREBP-2 transgenic mice: a new model for lipotoxicity. Journal of lipid research

543 2008; 49: 2524-2534. 2008/08/07. DOI: 10.1194/jlr.M800238-JLR200.

- 544 33. Laflamme D. Development and validation of a body condition score system for cats: a
- 545 clinical tool. Feline practice 1997; 25: 13-18.
- 546 34. Hoenig M, Thomaseth K, Waldron M, et al. Fatty acid turnover, substrate oxidation, and

547 heat production in lean and obese cats during the euglycemic hyperinsulinemic clamp. Domestic

- 548 *animal endocrinology* 2007; 32: 329-338. 2006/05/12. DOI: 10.1016/j.domaniend.2006.04.003.
- 549 35. Lund EM, Armstrong P, Kirk CA, et al. Prevalence and risk factors for obesity in adult cats

from private US veterinary practices. *Intern J Appl Res Vet Med* 2005; 3: 88-96.

- 551 36. Fettman MJ, Stanton CA, Banks LL, et al. Effects of neutering on bodyweight, metabolic rate
- and glucose tolerance of domestic cats. *Research in veterinary science* 1997; 62: 131-136. 1997/03/01.
- 553 37. Wara A and Datz C. Feline nutrition: The association between spaying or neutering and

weight gain. *Advances in Small Animal Medicine and Surgery* 2013; 26: 1-2.

- 555 38. Hewson-Hughes AK, Hewson-Hughes VL, Miller AT, et al. Geometric analysis of
- 556 macronutrient selection in the adult domestic cat, Felis catus. The Journal of experimental biology
- 557 2011; 214: 1039-1051. 2011/02/25. DOI: 10.1242/jeb.049429.
- 558 39. Laflamme DP. Companion Animals Symposium: Obesity in dogs and cats: What is wrong
- 559 with being fat? Journal of animal science 2012; 90: 1653-1662. 2011/10/11. DOI: 10.2527/jas.2011-
- 560 4571.
- 561 40. Hoenig M, Wilkins C, Holson JC, et al. Effects of obesity on lipid profiles in neutered male
- and female cats. *American journal of veterinary research* 2003; 64: 299-303. 2003/03/29.

- 563 41. Musco N, Calabro S, Tudisco R, et al. Diet effect on short- and long-term glycaemic response
- **in adult healthy cats**. *Veterinaria italiana* 2017; 53: 141-145. 2017/07/05. DOI:
- 565 10.12834/VetIt.57.166.3.
- 566 42. Eisert R. Hypercarnivory and the brain: protein requirements of cats reconsidered.
- 567 Journal of comparative physiology B, Biochemical, systemic, and environmental physiology 2011; 181: 1-
- 568 17. 2010/11/20. DOI: 10.1007/s00360-010-0528-0.
- 569 43. Colagiuri S and Brand Miller J. The 'carnivore connection'--evolutionary aspects of insulin
- 570 **resistance**. *European journal of clinical nutrition* 2002; 56 Suppl 1: S30-35. 2002/04/20. DOI:
- 571 10.1038/sj.ejcn.1601351.
- 572 44. Costacou T and Mayer-Davis EJ. Nutrition and prevention of type 2 diabetes. Annual review
- 573 *of nutrition* 2003; 23: 147-170. 2003/03/11. DOI: 10.1146/annurev.nutr.23.011702.073027.
- 574 45. Russell K, Sabin R, Holt S, et al. Influence of feeding regimen on body condition in the cat.
- 575 *The Journal of small animal practice* 2000; 41: 12-17. 2000/03/14.
- 576
- 577
- 578
- 579
- 580
- 581
- 582
- 583
- 584
- 585
- 586

587		
588		
589		
590		
591		
592		
593		
594		
595		
596		
597		