Review Article

ASTROBIOLOGY Volume 22, Number 2, 2022 Mary Ann Liebert, Inc. DOI: 10.1089/ast.2021.0066

> Open camera or QR reader and scan code to access this article and other resources online.



Out of Thin Air? Astrobiology and Atmospheric Chemotrophy

Don A. Cowan,¹ Belinda C. Ferrari,² and Christopher P. McKay³

Abstract

The emerging understanding of microbial trace gas chemotrophy as a metabolic strategy to support energy and carbon acquisition for microbial survival and growth has significant implications in the search for past, and even extant, life beyond Earth. The use of trace gases, including hydrogen and carbon monoxide as substrates for microbial oxidation, potentially offers a viable strategy with which to support life on planetary bodies that possess a suitable atmospheric composition, such as Mars and Titan. Here, we discuss the current state of knowledge of this process and explore its potential in the field of astrobiological exploration. Key Words: Astrobiology—Mars—Titan—Trace gas—Chemotrophy—Redox couple. Astrobiology 22, 225–232.

1. Introduction to Atmospheric Chemotrophy

TMOSPHERIC CHEMOSYNTHESIS, also termed trace gas A chemotrophy, is a set of metabolic processes in some microorganisms by which the oxidation of atmospheric trace gases, particularly molecular hydrogen (H_2) , methane (CH_4) , and carbon monoxide (CO), provide the metabolic energy required to drive carbon fixation via one of the most wellknown CO₂ assimilation processes, the Calvin-Benson-Bassham (CBB) cycle (Ji et al., 2017). Hydrogen, carbon monoxide, and methane are present in Earth's atmosphere at low concentrations, typically around 0.6, 0.1, and 1.85 ppm, respectively. A growing number of microbial taxa have been shown to possess the ability to oxidize these gases for energy generation through aerobic respiration, while others use methane and CO as the building blocks for life. Methanotrophy, where atmospheric methane supports both the carbon and energy requirements of select microbial taxa (Lau et al., 2015; Tveit et al., 2019), is probably the best known example of atmospheric chemotrophy. Interestingly, the thenhighly speculative concept that H₂ might act as an energy source that supports microbial survival has existed for more than three decades and was comprehensively reviewed by Morita over two decades ago (Morita, 2000).

In the cold desert soils of Eastern Antarctica, some of which lack photoautotrophs, microbially mediated atmospheric gas chemosynthesis, based specifically on assimilation of CO and H₂, is thought to support primary production. While it was first assumed that this "overlooked" primary production strategy was a unique and specialized process, there is a growing body of evidence that suggests trace gas assimilation is ubiquitous (Piché-Choquette and Constant, 2019; Bay et al., 2021), and atmospheric chemotrophy may be a generalist metabolic process in desert (and maybe other) soil microbial communities around the globe (Ray et al., 2020; Bay et al., 2021). Given that bacteria with the genetic capacity for trace gas assimilation are thought to literally "live on air," this discovery has significant implications for early life on Earth and in the search for biosignatures of extant or extinct life on other planets, particularly Mars.

2. The Genetics and Enzymology of Atmospheric Chemotrophy

The discovery of atmospheric (H₂/CO-dependent) chemotrophy as a primary production strategy was based on the application of genome-resolved metagenomics (Ji et al., 2017). This autotrophic process is proposed to involve three

¹Centre for Microbial Ecology and Genomics, Department of Biochemistry, Genetics and Microbiology, University of Pretoria, Pretoria,

South Africa. ²School of Biotechnology and Biomolecular Sciences, Australian Centre for Astrobiology, UNSW Sydney, Randwick, Australia. ³NASA Ames Research Center, Moffett Field, California, USA.

[©] Don A. Cowan et al., 2021; Published by Mary Ann Liebert, Inc. This Open Access article is distributed under the terms of the Creative Commons License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

core enzymes: hydrogenases, carbon monoxide dehydrogenases, and ribulose bisphosphate carboxylase (RuBisCO). Together, these enzymes act to acquire energy from trace gas oxidation and drive CO_2 assimilation (through the CBB cycle) to support cellular biosynthetic processes.

Interestingly, both autotrophic and mixotrophic lifestyles have been shown to be supported by trace gas oxidation, with the electrons produced by aerobic respiration used to assimilate CO and CO₂ via several pathways (King and Weber, 2007; Constant *et al.*, 2010; Greening *et al.*, 2014; Cordero *et al.*, 2019). However, trace gas autotrophy as a primary carbon fixation strategy has only been associated with the CBB cycle (Grostern and Alvarez-Cohen, 2013; Ji *et al.*, 2017; Tveit *et al.*, 2019).

Hydrogenases are metalloenzymes that are involved in the reversible reaction $(2H_2 + O_2 = 2H_2O)$ and include both assimilatory and synthetic enzymes, involved in hydrogen uptake and production, respectively. In soil, approximately 80% of H₂ uptake is driven by microbially mediated reactions that encompass two distinct processes (Constant et al., 2008). These biphasic processes include classical, lowaffinity reactions performed by Knallgas bacteria in the presence of high amounts of substrate (K_M H₂, ca. 1000 ppmv) and high-affinity reactions ($K_M H_2$, < 100 ppmv) that were first demonstrated in Streptomyces sp. PCB7 in 2008 (Schuler and Conrad, 1990; Constant et al., 2008). Subsequently, dose-response relationships have been observed in soil communities where elevated H₂ concentrations resulted in stimulation of low-affinity hydrogenase-expressing Knallgas bacteria and inhibition of high affinity H₂-oxidizers (Schuler and Conrad, 1990; Piché-Choquette et al., 2018; Piché-Choquette and Constant, 2019). Given the widespread abundance of type 1 [NiFe]-hydrogenases in oligotrophic environments, high-affinity oxidation appears to occur when substrates required for low-affinity hydrogenotrophy are limited, thus providing an alternative and stable energy source for cellular maintenance.

The newly discovered assimilatory type 1h [Ni-Fe]hydrogenases appear to be uniquely involved in scavenging of atmospheric H₂ for energy generation (Constant *et al.*, 2010; Greening *et al.*, 2014, 2015, 2016; Liot and Constant, 2016). Along with carbon monoxide dehydrogenases, these enzymes enable the use of H₂ and CO as electron donors for the aerobic respiratory chain, primarily to support cellular survival, but also in some cases growth (Jordaan *et al.*, 2020; Tveit *et al.*, 2021). To date, assimilatory hydrogenases have been identified in taxonomically diverse species of prokaryotes, with organisms harboring [NiFe]-hydrogenase encoding genes distributed across a significant number of soil taxa, currently spanning 36 bacterial and 6 archaeal phyla (Vignais and Billoud, 2007; Greening *et al.*, 2016).

The oxidation of atmospheric CO, catalyzed by type1 [MoCu] CO-dehydrogenases, also provides energy and carbon for aerobic carboxydotrophic bacteria to assimilate CO and/or CO₂ via RuBisCO, the primary step of the most well-known carbon fixation pathway, the CBB cycle (King and Weber, 2007). It is thought that atmospheric chemotrophic bacteria utilize a particular lineage of high substrate-affinity RuBisCO (type 1E). This enzyme remains relatively novel: first discovered in the *Mycobacterium* sp. strain JCI DSM 3803 (Park *et al.*, 2009), it has subsequently been linked to the soil Actinobacterium *Pseudonocardia dioxivorans* CB1190

during autotrophic growth (Grostern and Alvarez-Cohen, 2013) and to volcanic cave microbial communities on Mt Erebus, Antarctica (Tebo *et al.*, 2015). It has also been identified in polar soil mesocosms following hydrogen stimulation (Ji *et al.*, 2017; Ray *et al.*, 2020). Given the relatively recent discovery of atmospheric chemotrophy as a major survival and maintenance energy strategy in microbial communities that occupy hyper-oligotrophic soil niches (Leung

3. Extant Microbiology and the Ecological Importance of Atmospheric Chemotrophy

et al., 2020), pure culture studies are still required to vali-

date the proposed metabolic pathways and enzymes.

The enzymatic oxidation of gaseous hydrogen is a wellestablished process in terrestrial and marine microbial communities (Conrad, 1999; King, 2003). However, the critical ecological importance of this process was only recently highlighted with the discovery that trace gas chemotrophy is the dominant carbon and energy acquisition process in unique and environmentally extreme Antarctic soils, where microbial photoautotrophy (largely Cyanobacterial) is very limited or totally absent. This specialized process appears to be restricted to the ubiquitous Actinobacteriota and uncultured members within two candidate phyla: Ca. Dormibacterota and Ca. Eremiobacterota (previously known as AD3 and WPS-2, respectively) (Ji et al., 2017). Both candidate phyla are found in soil ecosystems but are usually rare, being present at abundances of <1% (Coveley et al., 2015; Ji et al., 2021). Surprisingly, however, in Robinson Ridge (East Antarctica) soils where atmospheric chemosynthesis was first identified, these candidate phyla dominated, representing up to 28% of the soil bacterial taxa detected (Ferrari et al., 2016; Ji et al., 2016).

In Antarctic desert soils, this process was implicated in the provision of maintenance energy in the survival of "dormant" microbial populations (Ji et al., 2017). Thermodynamic calculations based on published maintenance energy requirements for dormant cells (Conrad, 1999; Constant et al., 2010) suggest that atmospheric trace gas assimilation rates are compatible with minimum energy needs (Ji et al., 2017). More recently, hydrogenase genes (including those that encode the type 1h high-affinity assimilatory enzymes) have been found to be widespread in hot desert soils from around the world and associated with a wide range of taxa (Jordaan et al., 2020). In addition, key trace gas assimilating genes have been detected in desert soils that span the "three poles" (Ray et al., 2020), and, most recently, rapid trace gas oxidation rates have been quantitated in soils from across Australia (Jordaan et al., 2020; Bay et al., 2021). The suggestion that H_2/CO trace gas-driven autotrophy is a globally widespread process in soils (Ray et al., 2020; Bay et al., 2021), as found for atmospheric methanotrophy (Tveit *et al.*, 2019), has contradicted earlier suggestions that this microbially mediated strategy was restricted to extreme soil habitats where the capacity for phototrophy was limited (Ji et al., 2017).

Thermodynamic modeling continues to support the hypothesis that the energy derived from trace gas oxidation can theoretically support heterotrophic microbial communities at times of organic-carbon limitation (Bay *et al.*, 2021). Very recently, it has been discovered that the methylotrophic bacterium *Methylocapsa gorgona* MG08 is able to simultaneously oxidize all three common atmospheric trace gases (CH₄, CO, and H₂) to support aerobic growth (Tveit *et al.*, 2021).

Covariation between high-affinity H₂ and CO oxidation rates along a H₂ gradient, combined with field observations of the key enzymes responsible in a range of terrestrial ecosystems (Piché-Choquette et al., 2018), including volcanic deposits (Lynch et al., 2014) and Antarctic desert soils (Ji et al., 2017), suggests that atmospheric chemotrophs share similar environmental niches. Quantitative phylogenetic data also suggest that these organisms often belong to the rare biosphere, that is, low abundance (<1%) community members (although some members of the Actinobacteriota, the dominant taxa in many soils, harbor the potential for trace gas chemotrophy). However, these taxa appear to provide unique and critical services in oligotrophic soil ecosystems (Woodcroft et al., 2018, Brewer et al., 2019; Ji et al., 2021) and have now been detected in a growing number of oligotrophic and climatically extreme soil ecosystems, including the McMurdo Dry Valleys, the Tibetan plateau (Ray et al., 2020), and Mars analog sites such as the Andean Altiplano saltpans and the Atacama Desert (Lynch et al., 2014).

There is an emerging belief that microbial chemotrophy and photoautotrophy are likely to coexist. One possibility is that trace gas chemotrophy supports carbon and energy acquisition in some ecosystems but supplements alternative sources in others (Ji *et al.*, 2017; Ray *et al.*, 2020). However, it has recently been demonstrated that, within the yet-to-be cultured phylum *Ca*. Eremiobacterota, the order Baltobacterales contains members genetically capable of both anoxygenic photosynthesis and trace gas chemotrophy (Ward *et al.*, 2019; Ji *et al.*, 2021). It seems likely, therefore, that trace gas chemotrophy contributes to the carbon and energy assimilation in all but the most specialized microbial communities, although the relative contributions of trace gas chemotrophy and photoautotrophy in terrestrial and marine habitats have yet to be quantitated.

One of the characteristics of metabolic hydrogen oxidation is hydrogenesis; that is, it is water-generating. The ecological relevance of this physiological property is unknown, although it is unlikely to be of any significance in watersufficient environments. However, in arid and hyperarid soils, which constitute a significant proportion of Earth's land surface area (United Nations Decade Report, 2020), metabolic hydrogenesis may play a significant role (Ortiz *et al.*, 2021; in press). Specialized sublithic and endolithic habitats are the dominant microbial biotopes in many hot and cold deserts (Cary et al., 2010; Cowan et al., 2020), where the microbial communities are typically embedded in extracellular polysaccharide (EPS) matrices (de los Ríos et al., 2014) that both acquire and retain water (through the hygroscopic properties of the matrix: Roberson and Firestone, 1992; Costa et al., 2018). While no quantitative estimates currently exist for in situ metabolic hydrogenesis in these specialized niche habitats, we suggest that this process may make a significant contribution to water availability in these communities and, therefore, to water activity and metabolic capacity.

4. Planetary Atmospheres and Gas Evolution

By analogy with deserts on Earth, it has been suggested that dry surface soils on Mars could support biological consumption of atmospheric gases, based on the low levels of H₂, CO, O₂ in the martian atmosphere (Weiss *et al.*, 2000). Summers *et al.* (2002) modeled the release of CH₄ from the surface of Mars assuming H₂ (4H₂ + CO₂=CH₄ + 2H₂O) or CO (3CO +2H₂O=CH₄ + 2CO₂) as energy sources. It is generally agreed that moisture availability in the martian subsurface would be the limiting factor for biological uptake (Weiss *et al.*, 2000).

The availability of water on Mars could be ameliorated in two ways: either by implicating saturated salt solutions that depress the freezing point of water or by considering ice as a source of water activity. King (2015) showed that atmospheric CO (at 290 ppm) could be assimilated by halophilic microorganisms in saturated salt solutions, suggesting a possible basis for microbial activity in saturated brine. Brine flows that have been postulated as one explanation for the fluvial features, known as recurring slope lineae, have been frequently observed on Mars (Ojha *et al.*, 2014).

Trainer *et al.* (2019) reported on direct measurements of atmospheric gases over 3 martian years and found mixing ratios of O₂ to be 1.61 (\pm 0.09)×10⁻³ and CO=5.8 (\pm 0.8)×10⁻⁴. They found that the mixing ratio of O₂ showed an unexplained seasonal and inter-annual variability, by a factor of 2, ranging from 1.2×10⁻³ to 2.2×10⁻³. Krasnopolsky and Feldman (2001) reported the H₂ mixing ratio on Mars of 15±5 ppm in the lower atmosphere. In terms of absolute pressure, this would be 0.009 Pa, given the 610 Pa total pressure on Mars. In comparison, H₂ in Earth's atmosphere is 0.6 parts per million, equal to 0.06 Pa.

Reports of CH₄ in the martian atmosphere have varied, but the most detailed surface measurements (Webster *et al.*, 2018) indicate a roughly constant background of 0.4 ppbv with a seasonal variation that ranges from 0.24 to 0.65 ppbv. Webster *et al.* (2018) also reported temporary plumes with concentrations up to about 7 ppb. These values have, however, been called into question and may be due to outgassing by the Curiosity rover, since the Trace Gas Orbiter missions put an upper limit on CH₄ in the martian atmosphere at around 0.05 ppb (Korablev *et al.*, 2019). Due to its potentially long lifetime in the martian atmosphere (300 years), it has been suggested that CH₄ may be a useful indicator of biological activity (Summers *et al.*, 2002), as the stable gas would accumulate and diffuse globally.

The atmosphere of Titan, the largest of the saturnian moons, is dominated by N₂ with CH₄ at 5.6% and H₂ at 0.01% (Niemann *et al.*, 2010). In terms of chemical energy, C₂H₆ at 10 ppm and C₂H₂ at 1 ppm are of "biotic" interest (McKay and Smith, 2005). There is also a rich photochemistry of organic compounds in the upper atmosphere. However, the trace organics (with the exception of C₂H₄, at about 1 ppb at the surface) condense in Titan's 70 K tropopause (Fulchignoni *et al.*, 2005) and precipitate to the surface (*e.g.*, Vuitton *et al.*, 2019).

Titan is another interesting example of a location with surfaces that are at least occasionally moistened and in contact with an atmosphere where gases are out of equilibrium. The equatorial regions of Titan are dry but experience occasional rains (Graves *et al.*, 2008; Turtle *et al.*, 2011) that could wet the surface for several days (Williams *et al.*, 2012). The key factor is that, on Titan, the liquid is methane (and ethane), not water. The possibilities of life on Titan in liquid methane and ethane lakes and "rain"-soaked soils at

	TABLE 1. COMPARISON OF ENERGY THEEDS FOR METHANOGENESIS ON EARTH AND THAN		
Location	Reaction	Energy yields under local conditions	Reference
Titan Earth	$3H_2 + C_2H_2 = 2CH_4$ $4H_2 + CO_2 = CH_4 + 2H_2O$	111 kJ per mole H ₂ 24 kJ per mole H ₂	McKay and Smith, 2005 Sholes <i>et al.</i> , 2019

TABLE 1. COMPARISON OF ENERGY YIELDS FOR METHANOGENESIS ON EARTH AND TITAN

-180°C have recently been reviewed (McKay, 2016). The use of atmospheric gases as an energy source remains an intriguing possibility. Schulze-Makuch and Grinspoon (2005) and McKay and Smith (2005) suggested that H_2 and C_2H_2 could provide a redox couple to support biotic processes. In Titan's lower atmosphere, the mixing ratios of H_2 and C_2H_2 are 0.001 and 10 ppm, respectively, which implies a free energy of reaction of 111 kJ/mol of H_2 (McKay and Smith, 2005). A comparison of the free energy of this reaction on Titan with the free energy released by methanogenesis on Earth (Table 1) suggests that there is considerably more free energy available from the "Titan" reaction.

On Titan, the infrequent "rains" may serve the same role as a "water pulse" in deserts on Earth (Leung *et al.*, 2020), which trigger the measurable consumption of H_2 and C_2H_2 . The Dragonfly mission to Titan, scheduled for launch in 2027, will survey a range of sites on the surface of Titan. The payload includes a H_2 sensor to identify gas disequilibrium processes, as possible evidence of life in the liquid methane (Lorenz *et al.*, 2019).

While Earth, Mars, and Titan are the only worlds in the Solar System in which porous and occasionally moist soils are in contact with an atmosphere that contains redox couples, there are aquatic analogs in the outer Solar System. Jupiter's atmosphere contains a zone of clouds with water activity close to unity (Hallsworth *et al.*, 2021a). Cloud water droplets would be in contact with high H₂ levels and trace organics, which together, as on Titan, would provide a redox couple that could be exploited by biological systems. The saline ocean of Enceladus, one of Saturn's moons, contains low levels of H₂ and CO₂ and probably CO (Waite *et al.*, 2009, 2017), potentially available as biotic energy and carbon sources.

5. Physicochemical Limits for Life

The outer limits of life (the "biological envelope") have been a source of fascination for decades, particularly in the fields of extremophilic microbiology and astrobiology (Hallsworth et al., 2021b). A dramatic surge in studies of extremophilic microorganisms, since the 1970s, has led to a nearly stable definition of the biological envelope, where the outer "life" limits of the most dominant abiotic parameters (temperature, pH, radiation) are defined by the most extreme of Earth's extremophilic microorganisms (Harrison et al., 2013; von Hegner, 2020). Significant unknowns remain; however, the longevity of microorganisms under starvation conditions, desiccation (anhydrobiosis), frozen in permafrost (cryobiosis), in the deep subterranean biosphere or in hypersaline inclusions remains an issue of some uncertainty. These unresolved conundrums center on the mechanisms by which microorganisms can survive long periods of starvation limitation combined with one or more of the above states where, in the absence of basal cellular "maintenance" metabolism and repair processes, molecular degradation induced by background ionizing radiation should lead to cell death in much shorter time spans. The implication is, therefore, that viable cells in such specialized long-term survival environments must maintain some degree of metabolic activity capable of driving sufficient energy generation to support cellular maintenance and thus persist.

One of the concerning issues around this proposal has always been the concept that dormant cells are essentially ametabolic, such as in the state of anhydrobiosis (*e.g.*, Keilin, 1959). However, using both traditional (*e.g.*, radiolabeling: Cowan *et al.*, 1979) and modern molecular (*e.g.*, metatranscriptomics: León-Sobrino *et al.*, 2019) methods, researchers have come to perceive that dormant cells retain a low level of metabolic activity and persist. For example, it has recently been observed that some microbial taxa in hyperarid soils continue to express genes encoding nitrogen acquisition enzymes (particularly nitrate reductases: León-Sobrino *et al.*, 2019).

The other issue of concern is that continued metabolism requires a continuous supply of reduced substrate, and long-term utilization of oxidizable organic substrates is a non-equilibrium process; that is, organic substrates in the cellular microenvironment will be depleted. The implication of gas-dependent chemotrophy effectively resolves this dilemma, in that the high diffusability of atmospheric gasses and the low activation energy requirements are likely to continually replenish the reduced substrate supply (Piché-Choquette et al., 2018). We therefore suggest that trace gas chemotrophy, particularly H₂ assimilation, may play a critically important role in microbial longevity and survival in extremely oligotrophic, cold, and/or desiccated environments (such as might be found in a suitable astrobiological context). Hydrogen is a common, if low, concentration constituent of many planetary atmospheres and, in particular, is generated abiotically by the radiolysis of water in the presence of various minerals (LaVerne and Tonnies, 2003).

6. How Atmospheric Chemotrophy Might Support Life on Mars and Other Planetary Bodies

A long-standing conundrum in the search for extant life or for biomarkers of extinct life on exoplanets, particularly Mars, is what metabolic processes putative martian microbes could employ to obtain the energy required for cell maintenance and growth¹. The consumption of atmospheric

¹https://mepag.jpl.nasa.gov/meeting/2020-04/whitetopics/White Paper_07_Garvin.TraceGas.pdf

ASTROBIOLOGY AND ATMOSPHERIC CHEMOTROPHY

 H_2 , CO, and CO₂ by bacterial chemoautotrophs offers a versatile, minimalistic carbon acquisition strategy that offers new possibilities in the search for extraterrestrial life. The genetic determinants of this metabolic strategy have already been detected in Mars analog sites, which include the Andean Altiplano saltpans, the Atacama Desert (Lynch *et al.*, 2014), and mineral soils from both the Antarctic and high Arctic (Ray *et al.*, 2020).

The metabolic pathways for oxidation of CO and H₂ are considered some of the most ancient evolutionary processes thought to exist on Earth (Sholes et al., 2019), with prebiotic chemistry models predicting that the atmosphere on early Earth was rich in reducing gases (Tian et al., 2005; Piché-Choquette and Constant, 2019; Zahnle et al., 2020). The presence of the trace gases CO and H₂ in the martian atmosphere (Krasnopolsky and Feldman, 2001; Trainer et al., 2019) represents a potentially untapped source of free energy, much like that on Earth, which suggests that subsurface extant microbes should be capable of using trace gases as energy and carbon sources (Sholes et al., 2019). Using a one-dimensional photochemical model based on the free energy available from trace gases and predicting gas flux at the subsurface, the authors estimate maximum metabolic activity on the martian surface and calculate that approximately 10^{27} cells, equating to $2-40 \times 10^{-5}$ of Earth's biomass, could be supported through chemoautotrophic processes.

In the past, based on the assumption that the martian atmosphere was at equilibrium, it has been argued that extant life (essentially disequilibrium processes) could not be present (Lovelock, 1975, 1988). However, in recent years it has been reported that the martian atmosphere has the second largest thermodynamic disequilibrium after that of Earth, with approximately 136 J/mol of global atmospheric free energy (Krissansen-Totton *et al.*, 2016). This potentially available energy is linked to the redox pairs $CO-O_2$, O_2-CH_4 and, to a lesser extent, H_2-O_2 (Sholes *et al.*, 2019).

To some extent, the discovery that trace gas chemotrophy is probably a widespread metabolic process in Earth's terrestrial pan-microbiome offers a new, or at least stronger, paradigm for microbial survival on bodies beyond Earth, particularly Mars and Titan (Fig. 1). In particular, hydrogenotrophy offers exciting opportunities for the long-term energy sustenance of putative subsurface microbiomes. On Mars, the abiotic generation of this high-energy substrate, driven by radioautolysis over geological time, could provide a mechanism for the energetic support of evolutionary processes and for the continued supply of metabolic water, potentially facilitating the selection, adaptation, and survival of martian soil microorganisms on the "dying planet." On Titan, atmospheric photolysis produces redox couples that could support metabolism in exotic life-forms surviving in liquid hydrocarbons.

7. Signatures for Life Based on Signatures of Trace Gas Chemotrophs

The implications of trace gas chemotrophy as a minimalistic mode of carbon fixation are substantial, with the search for life on other planets now prioritizing H_2 and CO gas (Corenblit *et al.*, 2019; Sholes *et al.*, 2019). While a

> Occasional hydrocarbon



Iquid precipitation H₂ C₂H₂, C₂H₆ CH₄ Energy assimilation by H₂ Trace Gas Chemotrophy? Sub-surface microbiome?

FIG. 1. A model for the possible role of trace gas chemotrophy in the survival of soil microbiomes on Mars and Titan.

major research focus is on the development of biosignatures of H₂ and CO oxidation, there is also the prospect of exploiting redox pairs (CO-O₂ and H₂-O₂) as antibiosignatures, in the case that extant life does not exist (Sholes *et al.*, 2019). Experimental validation of trace gas carbon fixation will undoubtedly aid future predictions for life on exoplanets, particularly H₂ and CO consumption in the martian atmosphere.

Acknowledgments

The authors wish to thank their respective institutions (University of Pretoria, Pretoria, South Africa; University of New South Wales, Sydney, Australia; NASA Ames, La Jolla, USA) for the continued support of their research activities. B. Ferrari was supported through an Australian Research Council Future Fellowship (FT170100341).

References

- Bay SK, Dong X, Bradley JA, et al. (2021) Trace gas oxidizers are widespread and active members of soil microbial communities. Nat Microbiol 6:246–256.
- Brewer TE, Aronson EL, Arogyaswamy K, *et al.* (2019) Ecological and genomic attributes of novel bacterial taxa that thrive in subsurface soil horizons. *mBio* 10, doi:10.1128/ mBio.01318-19.
- Cary SC, McDonald I, Barrett JE, *et al.* (2010) On the rocks: the microbiology of Antarctic Dry Valley soils. *Nat Microbiol Rev* 8:129–138.
- Conrad R (1999) Soil microorganisms oxidizing atmospheric trace gases (CH₄, CO, H₂, NO). *Indian J Microbiol* 39:193– 203.
- Constant P, Poissant L, and Villemur R (2008) Isolation of *Streptomyces* sp. PCB7, the first microorganism demonstrating high-affinity uptake of tropospheric H₂. *ISME J* 2:1066–1076.
- Constant P, Chowdhury SP, Pratscher J, *et al.* (2010) Streptomycetes contributing to atmospheric molecular hydrogen soil uptake are widespread and encode a putative high-affinity [NiFe]-hydrogenase. *Environ Microbiol* 12:821– 829.
- Cordero PRF, Bayly K, Leung PM, *et al.* (2019) Atmospheric carbon monoxide oxidation is a widespread mechanism supporting microbial survival. *ISME J* 13:2868–2881.
- Corenblit D, Darrozes J, Julien F, *et al.* (2019) The search for a signature of life on Mars: a biogeomorphological approach. *Astrobiology* 19:1279–1291.
- Costa OYA, Raaijmakers JM, and Kuramae EE (2018) Microbial extracellular polymeric substances: ecological function and impact on soil aggregation. *Front Microbiol* 9, doi: 10.3389/fmicb.2018.01636.
- Coveley S, Elshahed MS, and Toussef NH (2015) Response of the rare biosphere to environmental stressors in a highly diverse ecosystem. *Peer J* 3, doi:10.7717/peerj.1182.
- Cowan DA, Green TGA, and Wilson AT (1979) Lichen metabolism. 1. The use of tritium labelled water in studies of anhydrobiotic metabolism in *Ramalina celastri* and *Peltigera polydactyla*. *New Phytol* 82:489–503.
- Cowan DA, Hopkins DW, Jones BE, *et al.* (2020) Microbiomics of Namib Desert habitats. *Extremophiles* 24: 17–29.

- de los Ríos A, Cary C, and Cowan DA (2014) The spatial structures of hypolithic communities in the Dry Valleys of East Antarctica. *Polar Biol* 37:1823–1833.
- Ferrari BC, Bissett A, Snape I, et al. (2016) Geological connectivity drives microbial community structure and connectivity in polar, terrestrial ecosystems. *Environ Microbiol* 18: 1834–1849.
- Fulchignoni M, Ferri F, Angrilli F, et al. (2005) In situ measurements of the physical characteristics of Titan's environment. Nature 438:785–791.
- Graves SDB, McKay CP, Griffith CA, *et al.* (2008) Rain and hail can reach the surface of Titan. *Planet Space Sci* 56:346–357.
- Greening C, Berney M, Hards K, et al. (2014) A soil actinobacterium scavenges atmospheric H₂ using two membraneassociated, oxygen-dependent [NiFe] hydrogenases. Proc Natl Acad Sci USA 111:4257–4261.
- Greening C, Constant P, Hards K, *et al.* (2015) Atmospheric hydrogen scavenging: from enzymes to ecosystems. *Appl Environ Microbiol* 81:1190–1199.
- Greening C, Biswas A, Carere CR, *et al.* (2016) Genomic and metagenomic surveys of hydrogenase distribution indicate H₂ is a widely utilised energy source for microbial growth and survival. *ISME J* 10:761–777.
- Grostern A and Alvarez-Cohen L (2013) RubisCO-based CO₂ fixation and C1 metabolism in the actinobacterium *Pseudonocardia dioxanivorans* CB1190. *Environ Microbiol* 15: 3040–3053.
- Hallsworth JE, Koop T, Dallas TD, *et al.* (2021a) Water activity in Venus' uninhabitable clouds and other planetary atmospheres. *Nat Astron* 5:665–675.
- Hallsworth JE, Mancinelli RL, Conley CA, *et al.* (2021b) Astrobiology of life on Earth. *Environ Microbiol* 23:3335– 3344.
- Harrison JP, Gheeraert N, Tsigelnitskiy D, et al. (2013) The limits for life under multiple extremes. Trends Microbiol 21: 204–212.
- Ji M, van Dorst J, Bissett A, et al. (2016) Microbial diversity at Mitchell Peninsula, Eastern Antarctica: a potential biodiversity "hotspot." Polar Biol 39:237–249.
- Ji M, Greening C, Vanwonterghem I, et al. (2017) Atmospheric trace gases support primary production in Antarctic desert surface soil. *Nature* 552:400–403.
- Ji M, Williams TJ, Montgomery K, *et al.* (2021) *Candidatus* Eremiobacterota, a metabolically and phylogenetically diverse terrestrial phylum with acid-tolerant adaptations. *ISME J* 15:2692–2707.
- Jordaan K, Lappan R, Dong X, et al. (2020) Hydrogenoxidizing bacteria are abundant in hot desert soils and strongly stimulated by hydration. mSystems 5, doi:10.1128/ mSystems.01131-20.
- Keilin D (1959) The Leeuwenhoek lecture—the problem of anabiosis or latent life: history and current concept. *Proc R Soc Lond B Biol Sci* 150:149–191.
- King GM (2003) Contributions of atmospheric CO and hydrogen uptake to microbial dynamics on recent Hawaiian volcanic deposits. *Appl Environ Microbiol* 69:4067–4075.
- King GM (2015) Carbon monoxide as a metabolic energy source for extremely halophilic microbes: implications for microbial activity in Mars regolith. *Proc Natl Acad Sci USA* 112:4465–4470.
- King GM and Weber CF (2007) Distribution, diversity and ecology of aerobic CO-oxidizing bacteria. *Nat Rev Microbiol* 5:107–118.

ASTROBIOLOGY AND ATMOSPHERIC CHEMOTROPHY

- Korablev O, Vandaele AC, Montmessin F, *et al.* (2019) No detection of methane on Mars from early ExoMars Trace Gas Orbiter observations. *Nature* 568:517–520.
- Krasnopolsky VA and Feldman PD (2001) Detection of molecular hydrogen in the atmosphere of Mars. *Science* 294: 1914–1917.
- Krissansen-Totton J, Bergsman DS, and Catling DC (2016) On detecting biospheres from chemical thermodynamic disequilibrium in planetary atmospheres. *Astrobiology* 16:39–67.
- Lau MCY, Stackhouse BT, Layton AC, et al. (2015) An active atmospheric methane sink in high Arctic mineral cryosols. *ISME J* 9:1880–1891.
- LaVerne JA and Tonnies SE (2003) H₂ production in the radiolysis of aqueous SiO₂ suspensions and slurries. *J Phys Chem B* 107:7277–7280.
- León-Sobrino C, Ramond J-B, Maggs-Kölling G, et al. (2019) Nutrient acquisition, rather than stress response over diel cycles, drives microbial transcription in a hyper-arid Namib Desert soil. Front Microbiol 10, doi:10.3389/fmicb.2019 .01054.
- Leung PM, Bay S, Meier D, *et al.* (2020) Energetic and trophic basis of microbial persistence in desert ecosystems. *mSystems* 5, doi:10.1128/mSystems.00495-19.
- Liot Q and Constant P (2016) Breathing air to save energy new insights into the ecophysiological role of high-affinity [NiFe]-hydrogenase in *Streptomyces avermitilis*. *MicrobiologyOpen* 5:47–59.
- Lorenz RD, Imanaka H, McKay CP, *et al.* (2019) Hydrogen sensing in Titan's atmosphere: motivations and techniques. *Planet Space Sci* 174:1–7.
- Lovelock JE (1975) Thermodynamics and the recognition of alien biospheres. *Proc R Soc Lond B Biol Sci* 189:167–181.
- Lovelock J (1988) Ages of Gaia. Oxford University Press, Oxford, UK.
- Lynch RC, Darcy JL, Kane NC, et al. (2014) Metagenomic evidence for metabolism of trace atmospheric gases by highelevation desert actinobacteria. Front Microbiol 5, doi: 10.3389/fmicb.2014.00698.
- McKay CP (2016) Titan as the abode of life. *Life* 6, doi: 10.3390/life6010008.
- McKay CP and Smith HD (2005) Possibilities for methanogenic life in liquid methane on the surface of Titan. *Icarus* 178: 274–276.
- Morita RY (2000) Is H₂ the universal energy source for longterm survival? *Microb Ecol* 38:307–320.
- Niemann HB, Atreya SK, Demick JE, et al. (2010) Composition of Titan's lower atmosphere and simple surface volatiles as measured by the Cassini-Huygens probe gas chromatograph mass spectrometer experiment. J Geophys Res Planets 115, doi:10.1029/2010JE003659.
- Ojha L, McEwen A, Dundas C, *et al.* (2014) HiRISE observations of recurring slope lineae (RSL) during southern summer on Mars. *Icarus* 231:365–376.
- Ortiz M, Leung PM, Shelley G, *et al.* (2021) Multiple energy sources and metabolic strategies sustain microorganisms in Antarctic desert soils. *Proc Natl Acad Sci USA*, 118 (45): e2025322118.
- Park SW, Hwang EH, Jang HS, et al. (2009) Presence of duplicate genes encoding a phylogenetically new subgroup of form I ribulose 1,5-bisphosphate carboxylase/oxygenase in *Mycobacterium* sp. strain JC1 DSM 3803. *Res Microbiol* 160: 159–165.
- Piché-Choquette S and Constant P (2019) Molecular hydrogen, a neglected key driver of soil biogeochemical pro-

cesses. Appl Environ Microbiol 285, doi:10.1128/AEM .02418-18.

- Piché-Choquette S, Khdhiri M, and Constant P (2018) Doseresponse relationships between environmentally relevant H₂ concentrations and the biological sinks of H₂, CH₄ and CO in soil. *Soil Biol Biochem* 123:190–199.
- Ray AE, Zhang E, Terauds A, *et al.* (2020) Soil microbiomes with the genetic capacity for atmospheric chemosynthesis are widespread across the poles and are associated with moisture, carbon, and nitrogen limitation. *Front Microbiol* 11, doi: 10.3389/fmicb.2020.01936.
- Roberson EB and Firestone MK (1992) Relationship between desiccation and exopolysaccharide production in a soil *Pseudomonas* sp. *Appl Environ Microbiol* 58:1284– 1291.
- Schuler S and Conrad R (1990) Soils contain two different activities for oxidation of hydrogen. *FEMS Microbiol Ecol* 6: 77–83
- Schulze-Makuch D and Grinspoon DH (2005) Biologically enhanced energy and carbon cycling on Titan? *Astrobiology* 5:560–567.
- Sholes SF, Krissansen-Totton J, and Catling DC (2019) A maximum subsurface biomass on Mars from untapped free energy: CO and H_2 as potential antibiosignatures. *Astrobiology* 19:655–668.
- Summers ME, Lieb BJ, Chapman E, et al. (2002) Atmospheric biomarkers of subsurface life on Mars. Geophys Res Lett 29: 24-1–24-4.
- Tebo BM, Davis RE, Anitori RP, *et al.* (2015) Microbial communities in dark oligotrophic volcanic ice cave ecosystems of Mt. Erebus, Antarctica. *Front Microbiol* 6, doi: 10.3389/fmicb.2015.00179.
- Tian F, Toon OB, Pavlov AA, *et al.* (2005) A hydrogen-rich early Earth atmosphere. *Science* 308:1014–1017.
- Trainer MG, Wong MH, McConnochie TH, et al. (2019) Seasonal variations in atmospheric composition as measured in Gale Crater, Mars. J Geophys Res Planets 124:3000– 3024.
- Turtle EP, Perry JE, Hayes AG, *et al.* (2011) Rapid and extensive surface changes near Titan's equator: evidence of April showers. *Science* 331:1414–1417.
- Tveit AT, Hestnes AG, Robinson SL, *et al.* (2019) Widespread soil bacterium that oxidizes atmospheric methane. *Proc Natl Acad Sci USA* 116:8515–8524.
- Tveit AT, Schmider T, Hestnes AG, *et al.* (2021) Simultaneous oxidation of atmospheric methane, carbon monoxide and hydrogen for bacterial growth. *Microorganisms* 9, doi: 10.3390/microorganisms9010153.
- United Nations Decade Report (2020) United Nations Decade for Deserts and the Fight Against Desertification. United Nations, New York. Available online at http://www.un.org/ en/events/desertification_decade/whynow.shtml
- Vignais PM and Billoud B (2007) Occurrence, classification, and biological function of hydrogenases: an overview. *Chem Rev* 107:4206–4272.
- von Hegner I (2020) Extremophiles: a special or general case in the search for extraterrestrial life? *Extremophiles* 24:167–175.
- Vuitton V, Yelle RV, Klippenstein SJ, *et al.* (2019) Simulating the density of organic species in the atmosphere of Titan with a coupled ion-neutral photochemical model. *Icarus* 324:120–197.
- Waite JH, Lewis WS, Magee BA, et al. (2009) Liquid water on Enceladus from observations of ammonia and 40 Ar in the plume. Nature 460:487–490.

- Waite JH, Glein CR, Perryman RS, et al. (2017) Cassini finds molecular hydrogen in the Enceladus plume: evidence for hydrothermal processes. *Science* 356:155–159.
- Ward LM, Cardona T, and Holland-Moritz H (2019) Evolutionary implications of anoxygenic phototrophy in the bacterial phylum *Candidatus* Eremiobacterota (WPS-2). *Front Microbiol* 10, doi:10.3389/fmicb.2019.01658.
- Webster CR, Mahaffy PR, Atreya SK, *et al.* (2018) Background levels of methane in Mars' atmosphere show strong seasonal variations. *Science* 360:1093–1096.
- Weiss BP, Yung YL, and Nealson KH (2000) Atmospheric energy for subsurface life on Mars? *Proc Natl Acad Sci USA* 97:1395–1399.
- Williams KE, McKay CP, and Persson F (2012) The surface energy balance at the Huygens landing site and the moist surface conditions on Titan. *Planet Space Sci* 60:376– 385.
- Woodcroft BJ, Singleton CM, Boyd JA, *et al.* (2018) Genomecentric view of carbon processing in thawing permafrost. *Nature* 560:49–54.
- Zahnle KJ, Lupu R, Catling DC, *et al.* (2020) Creation and evolution of impact-generated reduced atmospheres of early Earth. *Planetary Sci J* 1, doi:10.3847/PSJ/ab7e2c.

Address correspondence to: Don A. Cowan Centre for Microbial Ecology and Genomics Department of Biochemistry, Genetics and Microbiology University of Pretoria Building NW2, Room 3-12 Hatfield Campus Lynnwood Road Pretoria 0002 South Africa

E-mail: don.cowan@up.ac.za

Submitted 13 May 2021 Accepted 16 September 2021 Associate Editor: Lewis Dartnell

Abbreviations Used

CBB = Calvin-Benson-Bassham RuBisCO = ribulose bisphosphate carboxylase